#### Imidazo[1,2-a]quinoxalin-4-amines: A novel class of nonxanthine $A_1$ -adenosine receptor antagonists<sup>1</sup>

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**Abstract** – The syntheses and  $A_1$  adenosine receptor affinities of a number of imidazo[1,2-a]quinoxalin-4-amines are reported. Structure-activity relationships within the series and in comparison with other similar tricyclic nonxanthine adenosine antagonists are discussed, leading to a putative common binding mode of these nitrogen-containing heterocycles to  $A_1$  adenosine receptors. Secondary amino compounds displayed the best affinities toward  $A_1$  receptors, while the tertiary amines were almost devoid of activity, thus suggesting a crucial role for the hydrogen bond-forming 4-NH group. Remarkably higher potencies for 1-methyl and N-cyclopentyl derivatives were also found. 4-Cyclopentylamino-1-methylimidazo[1,2-a]quinoxaline (IRFI 165) is the most potent compound in this series, having  $K_i(A_1) = 7.9$  nM. It is also provided with a good  $A_1$  selectivity both versus  $A_{2a}$  and  $A_3$  subtypes and was selected for further pharmacological studies. © Elsevier, Paris

adenosine receptors / imidazo[1,2-a]quinoxaline / tricyclic heteroaromatic systems / nonxanthine A1-adenosine antagonists / IRFI 165

#### 1. Introduction

The nucleoside adenosine is involved in the regulation of several physiological functions which are mediated by the interaction with four distinct adenosine receptor (AR) subtypes, designated  $A_1$ ,  $A_{2a}$ ,  $A_{2b}$ , and  $A_3$  [1, 2, 3]. The search for subtype- and/or tissue-selective ligands of AR as novel pharmacological tools has been an active area of investigation, which has resulted in the synthesis and evaluation of thousands of new chemical entities as agonists, partial agonists, and antagonists [4-6]. The main potential therapeutic indications of  $A_1AR$  antagonists are for the treatment of cognitive deficits [7], renal failure [8], acute respiratory distress syndrome [9], cardiac arrhythmias [10] and asystolic cardiac arrest [11]. The naturally occurring xanthines, caffeine and theophylline, were the prototypic AR antagonists. The many attempts

to improve their potency and selectivity have resulted in the preparation of a large number of xanthine derivatives, and much is known now in terms of their structure—activity relationships (SAR) as well as of their pharmacological activity [12, 13]. In contrast, the structural requirements for receptor binding among the numerous different classes of nonxanthine AR antagonists are still not well defined [14].

In the course of a research program aimed to develop new centrally active A<sub>1</sub>AR antagonists [15], our efforts were directed toward the synthesis of tricyclic heteroaromatic compounds containing an exocyclic amine on a six-six-five ring system. Some of these structures (figure 1) have been described by other groups [16, 17, 18] and were reported to display AR antagonistic properties, with A<sub>1</sub>-selectivity being generally observed for N-cycloalkylamino derivatives. Among them, we focused our attention on the [1,2,4]triazolo[4,3-a]quinoxalin-4-amines described by Sarges et al. [19], who suggested that these compounds bind to AR by mimicking adenosine derivatives as shown in figure 2. From this overlap it appears that the 2-nitrogen atom may be unnecessary in order to achieve a good fitting and thus an optimal

<sup>&</sup>lt;sup>1</sup>Presented in part at the First Italian–Swiss Meeting on Medicinal Chemistry, Torino (Italy), 23–26 September 1997

<sup>\*</sup> Correspondence and reprints

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Figure 1. Basic structures of representative tricyclic nonxanthine adenosine antagonists.

interaction with the binding site of the receptor. Therefore, we synthesized a number of imidazo- and 1-methylimidazo [1,2-a] quinoxalin-4-amines (figure 3, compounds 1-26) and assayed them for  $A_1AR$  affinity.

#### 2. Chemistry

Monosubstitution of 2,3-dichloroquinoxaline (31a) with aminoacetaldehyde dimethylacetal or propargylamine gave the respective 2-(substituted) amino-3-chloroquinoxaline 27a and 28a, which, after treatment with strong mineral acid, underwent cyclization to imidazo- (29a) and 1-methylimidazo[1,2-a]quinoxalin-4(5H)-one (30a) in moderate to good yields (figure 4).

The above oxo heterocycles were converted to the final 4-amino derivatives via chlorination with POCl<sub>3</sub> or alternatively by an already described single-step silylation—amination procedure [20] using hexamethyldisilazane and the appropriate amine (figure 5).

Compounds 9–16 and 24–26 having chloro or fluoro substituents on the carbocyclic ring were prepared as described above from the corresponding 6-halo- and 6,7-dihalo-2,3-dichloroquinoxalines 31b–e, which were synthesized by a known route [21] from the appropriate o-phenylenediamine as illustrated in figure 6. The location of the halogen atom in the monochloro and monofluoro derivatives was unequivocally ascertained by 2-D NOE-ROESY experiments carried out on compounds 9

Figure 2. Putative binding mode of [1,2,4]triazolo[4,3-a]quinoxalin-4-amines: overlap with the agonist  $N^6$ -cyclopentyladenosine (drawn from [19]).

 $R_1 = H, CH_3;$ 

NR<sub>2</sub>R<sub>3</sub> = NH<sub>2</sub>, alkylamino, NH(CH<sub>2</sub>)<sub>2</sub>OH, cycloalkylamino, piperidino, morpholino, 4-methylpiperazino;

 $R_4, R_5 = H, Cl, F.$ 

**Figure 3.** Imidazo[1,2-a]quinoxalin-4-amines assayed for A<sub>1</sub>AR receptor affinity.

and 12 (as well as on the corresponding dihalogen derivatives 11 and 14 for comparison), which evidenced sharp negative crosspeaks due to a dipolar contact between the 1-H and the *meta*-coupled proton (9-H) adjacent to the halogen substituent.

#### 3. Pharmacology

The adenosine A<sub>1</sub>AR binding affinities of the imidazo[1,2-a]quinoxalin-4-amines were determined using standard radioligand binding assay procedures [22]. displacement of [<sup>3</sup>H]-1,3-dipropyl-8-Competitive cyclopentylxanthine ([3H]DPCPX) in rat brain membranes was used to determine a full concentration-inhibition curve for each compound. The binding potencies of the tested compounds, expressed as their  $K_i$  values, are listed in tables I and II. 4-Cyclopentylamino-1methylimidazo[1,2-a]quinoxaline (20, IRFI 165) was also assayed for its A2aAR and A3AR binding affinities against [3H]-2-[[4-(2-carboxyethyl)phenethyl]amino]-5'-(N-ethylcarboxamido)adenosine ([3H]CGS21680) using rat striatal membranes [23] and against [125I]-N6-(4amino-3-iodobenzyl)adenosine-5'-N-methyluronamide ([125I]AB-MECA) [24] using human recombinant (HEK-293) cells [25], respectively.

#### 4. Results

Table I shows that the 1-unsubstituted imidazo[1,2-a]quinoxalin-4-amines displayed  $K_i(A_1)$  values ranging from 23 nM to 5.6  $\mu$ M, with the exception of the tertiary amines (including N-piperidinyl, N-morpholinyl and N'-methyl-N-piperazinyl derivatives) 1-4 which were almost

**Figure 4**. (a)  $(H_3CO)_2CHCH_2NH_2$ , EtOH, reflux; (b) propargylamine, Et<sub>3</sub>N, EtOH, reflux; (c) 48% HBr (aq.), reflux; (d) conc.  $H_2SO_4$ , 90 °C.

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 $\textbf{Figure 5}. \ \ \textbf{(a) POCl}_3, \ C_6H_5N(CH_3)_2, \ \text{reflux; (b) NHR}_2R_3, \ \text{EtOH, reflux; (c) HMDS, (NH}_4)_2SO_4, \ \text{NHR}_2R_3, \ \text{reflux.}$ 

**Table I.** A<sub>1</sub>AR receptor affinities of the reported 1-(unsubstituted)-imidazo[1,2- $\alpha$ ]quinoxalin-4-amines.

Compound	$R_2$		R <sub>3</sub>	$R_4$	R <sub>5</sub>	$K_{i}(A_{1}), nM$
1	CH <sub>2</sub> CH <sub>3</sub>		CH <sub>2</sub> CH <sub>3</sub>	Н	Н	
2	2 3	-(CH <sub>2</sub> ) <sub>5</sub>	C112C113	H		≈ 25,000
3		-CH <sub>2</sub> CH <sub>2</sub> OCH <sub>2</sub> CH <sub>2</sub> -			H	≈ 64,000
4				H	Н	≈ 18,000
5	7.1	-CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>3</sub> )CH <sub>2</sub> CH <sub>2</sub> -		Н	Н	> 50,000
	H		$CH(CH_3)_2$	H	Н	1,430
0 -	H		c-C <sub>5</sub> H <sub>9</sub>	Н	H	175
/	Н		$c-C_6H_{11}$	Н	H	1,015
8	Н		CH <sub>2</sub> CH <sub>2</sub> OH	H	H	1,139
9	Н		$c-C_5H_9$	H	Cl	
10	H		c-C <sub>6</sub> H <sub>11</sub>	H	Cl	23.5
l1	Н		c-C <sub>5</sub> H <sub>9</sub>	Cl		394
12	Н		c-C <sub>5</sub> H <sub>9</sub>		Cl	26.5
13	H			H	F	84.5
14	H		c-C <sub>6</sub> H <sub>11</sub>	H	F	427
5	H		c-C <sub>5</sub> H <sub>9</sub>	F	F	298
			c-C <sub>6</sub> H <sub>11</sub>	Cl	Cl	5,601
16	Н		$CH(CH_2CH_3)_2$	H	Cl	301.5

**27b-e**:  $R = CH(OCH_3)_2$ 

28b-c:  $R = C \equiv CH$ 

$$X$$
 $NH_2$ 
 $R_4$ 
 $NH_2$ 
 $NH_2$ 
 $R_4$ 
 $NH_2$ 
 $NH_2$ 
 $R_4$ 
 $NH_2$ 
 $NH_2$ 
 $R_4$ 
 $NH_2$ 
 $NH$ 

Figure 6. (a) Diethyl oxalate, reflux; (b) POCl<sub>3</sub>, reflux.

 $\textbf{Table II.} \ A_1 AR \ receptor \ affinities \ of \ the \ reported \ 1-methylimidazo[1,2-a] quinoxalin-4-amines.$ 

**29b-e**:  $R_1 = H$ 

**30b-c**:  $R_1 = CH_3$ 

Compound	R <sub>2</sub>		$R_3$	$R_4$	R <sub>5</sub>	$K_{i}(A_{1}), nM$
17		-(CH <sub>2</sub> ) <sub>5</sub> -		Н	Н	> 30,000
18		-CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>3</sub> )CH <sub>2</sub> CH <sub>2</sub> -		Н	Н	> 30,000
19	Н	C112C1121 ((C113) C112C112	Н	H	Н	160.5
20 (IRFI 165)	H		c-C <sub>5</sub> H <sub>9</sub>	Н	Н	7.9
20 (IKIT 103) 21	H		c-C <sub>6</sub> H <sub>13</sub>	Н	H	114
22	H		CH <sub>2</sub> CH <sub>2</sub> OH	Н	Н	193.5
23	Н		CH(CH <sub>2</sub> CH <sub>3</sub> ) <sub>2</sub>	Н	Н	54
24	H		c-C <sub>5</sub> H <sub>9</sub>	Н	Cl	48
25	H		$c-C_6H_{11}$	Н	Cl	55.5
26	H		c-C <sub>5</sub> H <sub>9</sub>	Cl	Cl	77

devoid of affinity. It is noteworthy that the most potent compounds within this subgroup are the 4-cyclopentylamino derivatives, with compound 9 having the lowest  $K_i$  value. A significantly decreased affinity with respect to their lower homologues may be noticed for the cyclohexylamino derivatives (about 10-fold), while the open-chain analogue of 9 (16) also showed a reduced potency.

The 1-methylimidazo[1,2-a]quinoxalin-4-amines proved to be more potent than the corresponding 1-unsubstituted derivatives (table II), with the exception of compounds 24 and 26, both bearing a chloro substituent in the 8-position. 4-N-Piperidinyl and 4-N'-methyl-N-piperazinyl derivatives (17 and 18) showed again no activity at all, whereas the primary amine 19 exhibited a moderate affinity at A<sub>1</sub>AR. The most potent compound among the entire series is 20 (IRFI 165) which combined 4-cyclopentylamino and 1-methyl substitutions; it has a  $K_i(A_1)$  of 7.9 nM. This compound was also shown to be about 300-fold  $A_1$  selective versus  $A_{2a}AR$ , its  $K_i(A_{2a})$ being  $2.5~\mu M$ . In addition, a primary screen on human recombinant A<sub>3</sub>AR evidenced for IRFI 165 a 69% inhibition of the radioligand binding at 10 µM, thus suggesting a similarly reduced affinity also for this receptor subtype.

#### 5. Discussion

The structure-activity analysis of the tested compounds is consistent with the findings emerging from the study of similar tricyclic heteroaromatic systems such as those depicted in figure 1. In particular, the loss of potency for all the tertiary amines 1-4, 17 and 18 clearly suggests the occurrence of a strong interaction between the 4-NHR<sub>3</sub> substituent and a hydrogen-bond acceptor in the active binding site of the A<sub>1</sub>AR. Another common feature in the SAR patterns of these tricyclic systems is the increase in A<sub>1</sub> potency and selectivity seen with the N-cyclopentyl substitution at the exocyclic amine, and this finding is again confirmed by our results. Furthermore, the increase in A<sub>1</sub> affinity observed for 1-methyl derivatives in the present series parallels similar enhancements found with the introduction of lipophilic substituents at the C-1 position of [1,2,4]triazolo [4,3a]quinoxalin-4-amines [17] and at the C-2 position of 1H-imidazo[4,5-c]quinolin-4-amines [18]. In our series, however, a lack of additivity of the effects of 1-methyl and 8-chloro substituents may be noticed, suggesting that these nearby groups may occupy, at least in part, the same subregion of the active binding site. Alternatively, it may be argued that a limited overall lipophilicity in the

substituents at C-1, C-8 and, possibly, C-7 is required for an optimal interaction with the receptor.

The comparable A<sub>1</sub> binding potencies between the [1,2,4]triazolo[4,3-a]quinoxalin-4-amines described by Sarges et al. [19] and their 2-deaza analogues of the present study, as well as the similarities in the SAR of both series, show that the binding mode depicted in figure 2 may readily be shared by these tricyclic systems, thus confirming that the 2-nitrogen is unlikely to be crucially involved in the interaction with the receptor. It is noteworthy that the fitting shown in figure 2 has recently been supported by a theoretical SAR study indicating that this receptor-bound orientation give both electrostatic potential and shape similarities to a number of different classes of adenosine receptor ligands [26].

Thus, on the basis of the binding data and in close analogy with the tentative map of the binding site for the A<sub>1</sub> receptor proposed in the above-mentioned study, we were able to draw a putative pharmacophore accommodating 4-(cyclopentylamino)imidazo[1,2-a]quinoxalines (figure 7). Besides the H-bond acceptor and the H-bond donor respectively interacting with the 4-NHR and the N-3, which are likely to be essential for the binding, the presence of a cycloalkyl binding site with apparently strict steric requirements and an hydrophobic region having a limited degree of lipophilicity accommodating ability has also been suggested. In agreement with the previously reported models [26, 27], we tentatively propose for IRFI 165 the superimposition with the reference xanthine antagonist DPCPX illustrated in figure 8. In this fit the 4-NH of IRFI 165 overlaps with the N7-H of xanthines (which was shown also by our group to be equally essential for binding [28]), carbonyl oxygen in 6-position of xanthine occupies the same position as the N-3, the bridging nitrogen of the imidazoquinoxaline corresponds to the N-1 of DPCPX and, interestingly, the 1-methyl substituent is quite well superimposed with the N1-propyl of the xanthine.

The findings of the present study differ substantially from those of Colotta et al. [29], who assayed a limited number of imidazo[1,2-a]quinoxalines having 2-aryl substituents and a NH $_2$  group at position 4 and found a reduced A $_1$ AR affinity. However, we believe that the reason for this discrepancy has to be ascribed to the different nature of the substituents, particularly at the exocyclic amine.

The antagonistic character of the imidazo[1,2-a]quinoxalin-4-amines described in the present study may be easily understood from the close structural relationships with the known antagonists depicted in figure 1. Moreover, IRFI 165, as well as several of its analogues, showed a potent anti-immobility activity on

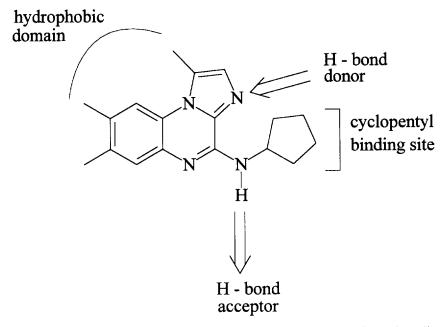


Figure 7. Suggested pharmacophore model accommodating 4-(cyclopentylamino)imidazo[1,2-a]quinoxalines.

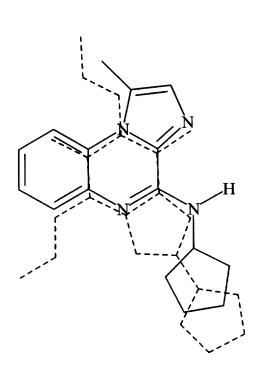


Figure 8. Proposed receptor-bound orientation of compound 20, fitted against 1,3-dipropyl-8-cyclopentylxanthine.

the Porsolt's forced swimming test in rodents [30], an effect which is likely to be mediated by an A<sub>1</sub>-antagonist induced enhancement of the release of various central neurotransmitters.

In conclusion, this study has provided further insights in the definition of the SAR of nonxanthine heteroaromatic A<sub>1</sub>AR antagonists, allowing a greater understanding of the structural requisites for maximizing A<sub>1</sub> affinity. In addition, a new potent and selective A<sub>1</sub>AR ligand (i.e. compound **20**, IRFI 165) has been identified and selected for further evaluation. The in vivo pharmacological characterization of IRFI 165 will be fully described elsewhere.

#### 6. Experimental protocols

#### 6.1. Chemical syntheses

Melting points were determined by differential scanning calorimetry (DSC) on a Perkin Elmer DSC 7 instrument. Microanalytical data were taken on a Carlo Erba 1106 analyzer or were obtained from Redox snc, Cologno M., Italy. Analyses indicated by the symbols of the elements were within ±0.4% of theoretical values. Infrared (IR) spectra were recorded on a Perkin Elmer 881 spectrophotometer as KBr pellets. <sup>1</sup>H-NMR spectra were obtained on a Varian EM 360L instrument or on a Bruker AM400 spectrometer using tetramethylsilane

(TMS,  $\delta$  0.00) or CHCl<sub>3</sub> contained in the deuterated solvent ( $\delta$  7.24) as the internal standard. Silica-gel plates (Merck F<sub>254</sub>) were used for analytical TLC. Chromatographic separations were performed on silica gel 60 (Merck, 230–400 mesh) in a preparative medium pressure liquid chromatographic apparatus (Gilson 305) equipped with UV (254 nm) detector (Gilson 112).

2,3,6-Trichloroquinoxaline (31b) was obtained from Acros Organics; all the other fine chemicals were from Aldrich.

#### 6.1.1. Imidazo[1,2-a]quinoxalin-4(5H)-ones (**29a-e**)

A mixture of 5.0 g (25 mmol) of 2,3-dichloro-quinoxaline (**31a**) and 5.5 mL (50 mmol) of aminoacetal-dehyde dimethyl acetal in 75 mL of absolute ethanol was refluxed for 4 h. Evaporation of the organic solvent under reduced pressure gave a residue which was dissolved in ethyl acetate. The organic solution was washed with water and brine, dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>) and evaporated at reduced pressure to afford a crude product which was purified by column chromatography (eluant: CH<sub>2</sub>Cl<sub>2</sub>), obtaining 5.0 g of 2-chloro-3-(2,2-dimethoxy-ethylamino)quinoxaline **27a** (IR: 3347, 2936, 1580, 1523, 1129 cm<sup>-1</sup>).

Compound **27a** (4.5 g, 16.8 mmol) was treated with 20 mL of 48% aqueous HBr and the mixture was refluxed for 4 h. Upon cooling the mixture was neutralized by aqueous NaOH and the resulting precipitate was filtered under suction and dried to obtain 3.2 g of imidazo[1,2-a]quinoxalin-4(5H)-one (**29a**) [31] which was recrystallized from water. M.p. 329.1 °C (DSC onset); IR (KBr): 3105 (NH), 1685 cm<sup>-1</sup> (C=O); <sup>1</sup>H-NMR (DMSO- $d_6$ ):  $\delta$  12.2 (1H, sb), 8.5 (1H, d, H-1, J = 2 Hz) 8.2–7.85 (1H, m), 7.5 (1H, d, H-2, J = 2 Hz), 7.4–7.05 (3H, m); UV (DMSO):  $\lambda_{\text{max}}$  = 284, 307, 320 nm. Anal.  $C_{10}H_7N_3O$  (C, H, N): calc. C 64.86, H 3.81, N 22.69%; found C 64.64, H 3.79, N 22.65%.

The following compounds were prepared in an analogous manner:

8-Chloroimidazo[1,2-a]quinoxalin-4(5H)-one (29b) (prepared from 2,3,6-trichloroquinoxaline (31b); recrystallized from DMF). M.p. > 350 °C (DSC); IR (KBr): 3144 (NH), 1724, 1684 cm<sup>-1</sup> (C=O); UV (DMSO):  $\lambda_{\text{max}}$  = 315, 327 nm. Anal. C<sub>10</sub>H<sub>6</sub>N<sub>3</sub>ClO (C, H, N): calc. C 54.69, H 2.75, N 19.13%; found C 54.37, H 2.75, N 19.11%.

7,8-Dichloroimidazo[1,2-a]quinoxalin-4(5H)-one (29c) [32] (prepared from 2,3,6,7-tetrachloroquinoxaline (31c); recrystallized from DMF). M.p. > 350 °C (DSC); IR (KBr): 3069 (NH), 3027 (CH), 1711 cm<sup>-1</sup> (C=O); <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>/DMSO- $d_6$ ):  $\delta$  12.25 (1H, sb), 8.70 (1H, d, H-1, J = 2 Hz), 8.63 (1H, s), 7.74 (1H, d,

H-1, J = 2 Hz), 7.66 (1H, s); UV (DMSO):  $\lambda_{\text{max}} = 289$ , 318, 331 nm. Anal.  $C_{10}H_5N_3Cl_2O$  (C, H, N): calc. C 47.27, H 1.98, N 16.54%; found C 47.37, H 2.02, N 16.45%.

8-Fluoroimidazo[1,2-a]quinoxalin-4(5H)-one (29d) (prepared from 2,3-dichloro-6-fluoroquinoxaline 31d; recrystallized from absolute ethanol). IR (KBr): 3131 (NH), 1698 cm<sup>-1</sup> (C=O); <sup>1</sup>H-NMR (DMSO- $d_6$ ):  $\delta$  12.2 (1H, sb), 8.7 (1H, d, H-1, J = 2 Hz), 8.3 (1H, dd, H-9,  $J_{o-HF}$  = 10 Hz), 7.8 (1H, d, H-1, J = 2 Hz), 7.6–7.2 (2H, m).

7,8-Difluoroimidazo[1,2-a]quinoxalin-4(5H)-one (29e) (prepared from 2,3-dichloro-6,7-difluoroquinoxaline (31e)). IR (KBr): 3127 (NH), 1683 cm $^{-1}$  (C=O);  $^{1}$ H-NMR (DMSO- $d_{6}$ ):  $\delta$  12.6 (1H, sb), 8.6 (1H, d, H-1, J = 2 Hz), 8.35–7.9 (2H, m), 7.7 (1H, dd, H-6,  $J_{o\text{-HF}}$  = 10 Hz,  $J_{m\text{-HF}}$  = 7 Hz).

## 6.1.2. 1-Methylmidazo[1,2-a]quinoxalin-4(5H)-ones (30a-c)

A mixture of 2,3-dichloroquinoxaline (10.0 g, 50 mmol), propargylamine (4.5 mL, 65 mmol), triethylamine (10.5 mL, 75 mmol) in 50 mL of absolute ethanol was heated at reflux for 4 h. Evaporation of the organic solvent under reduced pressure gave a residue which was purified by column chromatography (eluant:  $CH_2Cl_2$ ), obtaining 7.0 g of 2-chloro-3-(propargylamino) quinoxaline **28a** (IR: 3441, 3282, 1518 cm<sup>-1</sup>).

This compound (7.0 g, 32 mmol) was treated with 10 mL of concentrated  $H_2SO_4$  and the mixture was stirred at 90 °C for 1 h. Upon cooling the mixture was cautiously neutralized by aqueous NaOH and the resulting precipitate was filtered under suction, dried, decolorized and recrystallized from DMF to obtain 2.4 g of 1-methylimidazo[1,2-a]quinoxalin-4(5H)-one (30a). M.p. > 320 °C (DSC); IR (KBr): 3132 (NH), 2977 (CH), 1691 cm<sup>-1</sup> (C=O); <sup>1</sup>H-NMR (DMSO- $d_6$ ):  $\delta$  11.8 (1H, sb), 8.1–7.95 (1H, m), 7.4–7.1 (4H, m), 2.8 (3H, s); UV (DMSO):  $\lambda_{max} = 263$ , 292, 304 nm. Anal.  $C_{11}H_9N_3O$  (C, H, N): calc. C 66.32, H 4.55, N 21.09%; found C 66.22, H 4.62, N 20.89%.

The following compounds were prepared in an analogous manner:

8-Chloro-1-methylimidazo[1,2-a]quinoxalin-4(5H)-one (30b) (prepared from 2,3,6-trichloroquinoxaline 31b; recrystallized from DMF). IR (KBr): 3044 (CH),  $1689 \text{ cm}^{-1} \text{ (C=O)}$ ;  $^{1}\text{H-NMR} \text{ (DMSO-}d_{6})$ :  $\delta$  11.8 (1H, sb), 8.0 (1H, d, H-9, J = 2 Hz), 7.4–7.2 (3H, m), 2.85 (3H, s).

7,8-Dichloro-1-methylimidazo[1,2-a]quinoxalin-4(5H)-one (30c) (prepared from 2,3,6,7-tetrachloroquinoxaline **31c**; recrystallized from *N*,*N*-dimethylacetamide). IR (KBr):  $1684 \text{ cm}^{-1}$  (C=O);  $^{1}\text{H-NMR}$  (DMSO- $d_{6}$ ):  $\delta$  12.3 (1H, sb), 8.1 (1H, s), 7.6 (1H, s), 7.25 (1H, s), 2.8 (3H, s).

6.1.3. 4-Diethylaminoimidazo[1,2-a]quinoxaline (1)

Compound **29a** (0.47 g, 2.5 mmol) was treated with POCl<sub>3</sub> (5.6 mL) and *N*,*N*-dimethylaniline (0.42 mL) and the mixture was refluxed for 2 h. Excess reagents were removed under vacuum and the residue was taken up in CHCl<sub>3</sub> and washed with water until neutral reaction, dried, evaporated and recrystallized from *n*-hexane/CHCl<sub>3</sub>, affording 0.29 g of 4-chloroimidazo[1,2-*a*] quinoxaline. M.p. (DSC onset) 192.3 °C; IR (KBr): 3064, 1503 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  8.2 (1H, d, H-1, J = 2 Hz), 8.1–7.85 (1H, m), 7.8 (1H, d, H-2, J = 2 Hz), 7.75–7.4 (3H, m); UV (EtOH):  $\lambda_{\text{max}}$  = 247, 315, 330 nm. Anal. C<sub>10</sub>H<sub>6</sub>N<sub>3</sub>Cl (C, H, N): calc. C 58.98, H 2.97, N 20.64%; found C 58.75, H 2.98, N 20.57%.

A mixture of the above compound (2.8 g, 13.7 mmol) and diethylamine (9.8 mL, 94.7 mmol) in 40 mL of absolute ethanol was refluxed for 4 h. Evaporation of the organics under reduced pressure gave a residue which was taken up in CHCl<sub>3</sub> and washed with water and brine, dried, evaporated, purified by column chromatography (eluant: CH<sub>2</sub>Cl<sub>2</sub>) and recrystallized from *n*-hexane to obtain 1.3 g of the title compound. M.p. (DSC onset) 91.7 °C; IR (KBr): 2976, 1518, 1425 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  7.9 (1H, s), 7.7–7.4 (3H, m), 7.35–7.05 (2H, m), 4.15 (4H, q), 1.3 (6H, t); UV (EtOH):  $\lambda_{max}$  = 231, 250, 293, 305, 332, 348 nm. Anal. C<sub>14</sub>H<sub>16</sub>N<sub>4</sub> (C, H, N): calc. C 69.97, H 6.71, N 23.31%; found C 69.99, H 6.80, N 23.13%.

The following compounds were similarly prepared from the appropriate imidazo- (29a-e) or 1-methylimidazo [1,2-a] quinoxalin-4(5H)-one (30a-c).

6.1.4. 4-(N-piperidinyl)imidazo[1,2-a]quinoxaline (2) Recryst. solvent: ethanol/water. M.p. (DSC onset) 108.2 °C; IR (KBr): 3107, 2935, 1517 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  7.9 (1H, d, H-1, J = 2 Hz), 7.75–7.4 (3H, m), 7.4–7.1 (2H, m), 4.3 (4H, t), 1.9–1.6 (6H, m); UV (EtOH):  $\lambda_{\text{max}}$  = 231, 249, 293, 305, 333 nm. Anal.  $C_{15}H_{16}N_4$  (C, H, N): calc. C 71.40, H 6.39, N 22.20%; found C 71.38, H 6.63, N 22.61%.

6.1.5. 4-(N-morpholinyl)imidazo[1,2-a]quinoxaline (3) Recryst. solvent: absolute ethanol. M.p. (DSC onset) 142.8 °C; IR (KBr): 3016, 2962, 1517 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  8.0 (1H, s), 7.8–7.5 (3H, m), 7.5–7.2 (2H, m), 4.4 (4H, t), 3.9 (4H, t); UV (EtOH):  $\lambda_{\text{max}}$  = 230, 248, 292, 304, 330 nm. Anal. C<sub>14</sub>H<sub>14</sub>N<sub>4</sub>O (C, H, N): calc. C 66.13, H 5.55, N 22.03%; found C 65.43, H 5.47, N 22.25%.

## 6.1.6. 4-(N'-Methyl-N-piperazinyl)imidazo[1,2-a]qui-noxaline (4)

This compound was obtained as the dihydrochloride by treatment of the free base with ethanolic HCl. M.p. (DSC

onset) 305.6 °C; IR (KBr): 2697, 1560, 1508 cm<sup>-1</sup>; <sup>1</sup>H-NMR (DMSO- $d_6$ /CD<sub>3</sub>OD):  $\delta$  8.7 (1H, sb), 8.3–8.0 (1H, m), 7.8–7.35 (4H, m), 5.5 (4H, t), 3.7–3.3 (4H, m), 2.8 (3H, s); UV (EtOH):  $\lambda_{\text{max}}$  = 230, 291, 304, 320 nm. Anal. C<sub>15</sub>H<sub>17</sub>N<sub>5</sub>·2HCl (C, H, N): calc. C 52.95, H 5.63, N 20.58%; found C 52.81, H 5.78, N 20.13%.

6.1.7. 4-Isopropylaminoimidazo[1,2-a]quinoxaline (5) Recryst. solvent: *n*-hexane. M.p. (DSC onset) 102.7 °C; IR (KBr): 3230, 2966, 1559 cm<sup>-1</sup>; <sup>1</sup>H-NMR (DMSO- $d_6$ ):  $\delta$  8.55 (1H, s), 8.2–7.95 (1H, m), 7.6–7.4 (2H, m), 7.4–7.2 (3H, m), 4.5 (1H, m), 1.25 (6H, d); UV (EtOH):  $\lambda_{\text{max}}$  = 227, 244, 285, 297, 318, 332 nm. Anal.  $C_{13}H_{14}N_4$  (C, H, N): calc. C 69.00, H 6.23, N 24.76%; found C 69.17, H 6.70, N 25.05%.

6.1.8. 4-Cyclopentylaminoimidazo[1,2-a]quinoxaline (6) Recryst. solvent: DMF/water. M.p. (DSC onset) 114.3 °C; IR (KBr): 3419, 2962, 1543 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  7.9 (1H, s), 7.8–7.0 (5H, m), 6.2 (1H, d), 4.7 (1H, m), 2.3–2.05 (2H, m), 2.0–1.3 (6H, m); UV (EtOH):  $\lambda_{\text{max}} = 228, 244, 285, 297, 319, 333$  nm. Anal.  $C_{15}H_{16}N_4$  (C, H, N): calc. C 71.40, H 6.39, N 22.20%; found C 71.13, H 7.30, N 22.29%.

6.1.9. 4-Cyclohexylaminoimidazo[1,2-a]quinoxaline (7) Recryst. solvent: n-hexane/ethyl acetate. M.p. (DSC onset) 94.0 °C; IR (KBr): 3227, 2924, 1560 cm<sup>-1</sup>;  $^{1}$ H-NMR (CDCl<sub>3</sub>):  $\delta$  7.85 (1H, d, H-1, J = 2 Hz), 7.8–7.4 (3H, m), 7.4–7.1 (2H, m), 6.0 (1H, d), 4.2 (1H, m), 2.3–1.95 (2H, m), 1.95–0.9 (8H, m); UV (EtOH):  $\lambda_{max}$  = 225, 242, 283, 294, 316, 330 nm. Anal.  $C_{16}H_{18}N_4$  (C, H, N): calc. C 72.15, H 6.81, N 21.03%; found C 72.25, H 7.04, N 21.19%.

#### 6.1.10. 4-(2-Hydroxyethylamino)imidazo[1,2-a]quinoxaline (8)

Recryst. solvent: isopropyl acetate. M.p. (DSC onset) 150.8 °C; IR (KBr): 3312, 1598, 1564 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>/CD<sub>3</sub>OD):  $\delta$  8.2 (1H, s), 8.1–7.55 (2H, m), 7.55 (1H, s), 7.4–7.2 (2H, m), 3.8 (4H, s); UV (EtOH):  $\lambda_{\text{max}}$  = 227, 285, 297, 317, 330 nm. Anal.  $C_{12}H_{12}N_4O$  (C, H, N): calc. C 63.14, H 5.30, N 24.55%; found C 63.16, H 5.40, N 24.99%.

## 6.1.11. 8-Chloro-4-cyclopentylaminoimidazo[1,2-a] quinoxaline (9)

Recryst. solvent: *n*-hexane/ethyl acetate. M.p. (DSC onset) 140.1 °C; IR (KBr): 3401, 2955, 1554 cm<sup>-1</sup>; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.87 (1H, d, H-1, J = 2 Hz), 7.68–7.62 (2H, m), 7.53 (1H, d, H-2, J = 2 Hz), 7.35 (1H, dd, H-7), 6.10 (1H, s, N*H*), 4.68–4.58 (1H, m), 2.25–2.13 (2H, m), 1.85–1.55 (6H, m); UV (EtOH):  $\lambda_{\text{max}}$  = 229,

245, 289, 301, 326, 340 nm. Anal.  $C_{15}H_{15}ClN_4$  (C, H, N): calc. C 62.83, H 5.27, N 19.54%; found C 63.04, H 5.36, N 19.64%.

## 6.1.12. 8-Chloro-4-cyclohexylaminoimidazo[1,2-a] quinoxaline (10)

Recryst. solvent: ethanol/water. M.p. (DSC onset) 126.7 °C; IR (KBr): 3413, 2926, 1555 cm $^{-1}$ ;  $^{1}\text{H-NMR}$  (CDCl $_{3}$ ):  $\delta$  7.85 (1H, s), 7.7–7.2 (4H, m), 6.1 (1H, d), 4.2 (1H, m), 2.3–2.0 (2H, m), 2.0–1.2 (8H, m); UV (EtOH):  $\lambda_{\text{max}} = 229, 245, 289, 301, 326, 340 \text{ nm}.$  Anal.  $C_{16}H_{17}\text{ClN}_{4}$  (C, H, N): calc. C 63.89, H 5.70, N 18.63%; found C 64.07, H 5.79, N 18.80%.

## 6.1.13. 4-Cyclopentylamino-7,8-dichloroimidazo[1,2-a]quinoxaline (11)

Recryst. solvent: ethanol/water. M.p. (DSC onset) 139.4 °C; IR (KBr): 3247, 2961, 1589, 1556 cm<sup>-1</sup>; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.85 (1H, d, H-1, J = 2 Hz), 7.82 (1H, s, H-9), 7.73 (1H, s, H-6), 7.55 (1H, d, H-2, J = 2 Hz), 6.22 (1H, d, NH), 4.65–4.55 (1H, m), 2.22–2.12 (2H, m), 1.85–1.55 (6H, m); UV (EtOH):  $\lambda$ <sub>max</sub> = 235, 274, 292, 304, 330, 345 nm. Anal. C<sub>15</sub>H<sub>14</sub>Cl<sub>2</sub>N<sub>4</sub> (C, H, N): calc. C 56.09, H 4.39, N 17.44%; found C 56.13, H 4.41, N 17.52%.

## 6.1.14. 4-Cyclopentylamino-8-fluoroimidazo[1,2-a] quinoxaline (12)

Recryst. solvent: ethanol/water. M.p. (DSC onset) 85.7 °C; IR (KBr): 3255, 2964, 1551 cm<sup>-1</sup>; <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.83 (1H, d, H-1, J = 2 Hz), 7.68 (1H, dd, H-6,  $J_{m\text{-HF}}$  = 6 Hz), 7.54 (1H, d, H-2, J = 2 Hz), 7.34 (1H, dd, H-9,  $J_{o\text{-HF}}$  = 9 Hz), 7.17–7.11 (1H, m, H-7), 6.02 (1H, d, N*H*), 4.67–4.58 (1H, m), 2.23–2.12 (2H, m), 1.86–1.56 (6H, m); UV (EtOH):  $\lambda_{\text{max}}$  = 226, 268, 285, 296, 323, 336 nm. Anal.  $C_{15}H_{15}FN_4$  (C, H, N): calc. C 66.65, H 5.59, N 20.73%; found C 66.36, H 5.66, N 20.86%.

#### 6.1.15. 4-Cyclohexylamino-8-fluoroimidazo[1,2-a]quinoxaline (13)

Recryst. solvent: ethanol. M.p. (DSC onset) 157.6 °C; IR (KBr): 3415, 2927, 1556 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  7.8 (1H, dd, J = 2 Hz), 7.7–7.35 (1H, dd, J<sub>HF</sub> = 16 Hz), 7.55 (1H, d, J = 2 Hz), 7.55–7.25 (1H, dd, J<sub>HF</sub> = 16 Hz), 7.05 (1H, dd), 6.0 (1H, d), 4.25 (1H, m), 2.35–1.2 (10H, m); UV (EtOH):  $\lambda$ <sub>max</sub> = 226, 239, 285, 297, 323, 337 nm. Anal. C<sub>16</sub>H<sub>17</sub>FN<sub>4</sub> (C, H, N): calc. C 67.59, H 6.03, N 19.71%; found C 67.31, H 6.04, N 19.70%.

#### 6.1.16. 4-Cyclopentylamino-7,8-difluoroimidazo[1,2-a]quinoxaline (14)

Recryst. solvent: ethanol/water. M.p. (DSC onset) 146.0 °C; IR (KBr): 3263, 2955, 1554 cm $^{-1}$ ;  $^{1}\text{H-NMR}$  (400 MHz, CDCl $_{3}$ ):  $\delta$  7.78 (1H, d, H-1, J=2 Hz), 7.54 (1H, d, H-2, J=2 Hz), 7.51 (1H, dd, H-9,  $J_{o\text{-HF}}=10$  Hz,  $J_{m\text{-HF}}=7$  Hz), 7.44 (1H, dd, H-6,  $J_{o\text{-HF}}=10$  Hz,  $J_{m\text{-HF}}=7$  Hz), 6.12 (1H, d, NH), 4.65–4.55 (1H, m), 2.22–2.12 (2H, m), 1.85–1.05 (6H, m); UV (EtOH):  $\lambda_{\text{max}}=225, 241, 270, 295, 323, 337$  nm. Anal.  $C_{15}H_{14}F_{2}N_{4}$  (C, H, N): calc. C 62.49, H 4.89, N 19.43%; found C 62.46, H 5.03, N 19.65%.

#### 6.1.17. 4-Cyclohexylamino-7,8-dichloroimidazo[1,2-a]quinoxaline (15)

Recryst. solvent: ethanol/water. M.p. (DSC onset) 162.3 °C; IR (KBr): 3332, 2929, 1587, 1550 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  7.8 (2H, m), 7.7 (1H, s), 7.55 (1H, d, J = 2 Hz), 6.2 (1H, d), 4.4 (1H, m), 2.3–1.2 (10H, m); UV (EtOH):  $\lambda_{\text{max}}$  = 234, 274, 292, 304, 330, 345 nm. Anal.  $C_{16}H_{16}Cl_2N_4\cdot1/2H_2O$  (C, H, N): calc. C 55.82, H 4.98, N 16.28%; found C 55.83, H 5.00, N 16.31%.

## 6.1.18. 1-Methyl-4-(N-piperidinyl)imidazo[1,2-a]quino-xaline (17)

Recryst. solvent: ethanol/water. M.p. (DSC onset) 78.9 °C; IR (KBr): 3018, 2927, 1502 cm $^{-1}$ ;  $^{1}\text{H-NMR}$  (CDCl $_{3}$ ):  $\delta$  8.05 (1H, dd), 7.65 (1H, dd), 7.5–7.0 (3H, m), 4.25 (4H, t), 2.85 (3H, s), 1.9–1.6 (6H, m); UV (EtOH):  $\lambda_{\text{max}} = 249, 281, 297, 310, 329$  nm. Anal.  $C_{16}H_{18}N_{4}$  (C, H, N): calc. C 72.15, H 6.81, N 21.03%; found C 71.84, H 7.09, N 20.70%.

## 6.1.19. 1-Methyl-4-(N'-methyl-N-piperazinyl)imidazo [1,2-a]quinoxaline (18)

Recryst. solvent: *n*-hexane/isopropyl acetate. M.p. (DSC onset) 108.0 °C; IR (KBr): 2928, 1535, 1510 cm<sup>-1</sup>; 

<sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  8.1 (1H, dd), 7.7 (1H, dd), 7.4--7.0 (3H, m), 4.35 (4H, t), 2.85 (3H, s), 2.6 (4H, t), 2.3 (3H, s); UV (EtOH):  $\lambda_{max}$  = 228, 248, 279, 309, 325 nm. Anal. C<sub>16</sub>H<sub>19</sub>N<sub>5</sub> (C, H, N): calc. C 68.30, H 6.81, N 24.89%; found C 68.37, H 7.15, N 25.05%.

# 6.1.20. 1-Methylimidazo[1,2-a]quinoxalin-4-amine (19) Recryst. solvent: n-hexane/ethyl acetate. M.p. (DSC onset) 245.0 °C; IR (KBr): 3391, 1641, 1517 cm<sup>-1</sup>; $^{1}$ H-NMR (400 MHz, CDCl<sub>3</sub>/DMSO- $d_6$ ): $\delta$ 8.35 (1H, dd), 7.87 (1H, dd), 7.66 (1H, s), 7.65–7.56 (2H, m), 3.03 (3H, s); UV (EtOH): $\lambda_{\text{max}} = 258$ , 268, 301, 313 nm. Anal. $C_{11}H_{10}N_4$ (C, H, N): calc. C 66.65, H 5.08, N 28.26%; found C 66.56, H 5.10, N 28.21%.

## 6.1.21. 4-(2-Hydroxyethylamino)-1-methylimidazo [1,2-a]quinoxaline (22)

Recryst. solvent: ethanol/water. M.p. (DSC onset) 173.4 °C; IR (KBr): 3415, 3217, 1558 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  8.0 (1H, dd), 7.7–7.2 (3H, m), 7.2 (1H, s), 6.6 (1H, m), 5.3 (1H, sb), 3.85 (4H, m), 2.8 (3H, s); UV (EtOH):  $\lambda_{max}$  = 225, 242, 271, 301, 314, 327 nm. Anal.  $C_{13}H_{14}N_4O$  (C, H, N): calc. C 64.45, H 5.82, N 23.12%; found C 64.29, H 5.91, N 23.13%.

## 6.1.22. 4-(1-Ethylpropylamino)-1-methylimidazo[1,2-a]quinoxaline (23)

Recryst. solvent: *n*-hexane. M.p. (DSC onset) 75.9 °C; IR (KBr): 3231, 2965, 1546 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  8.0 (1H, dd), 7.7 (1H, dd), 7.4–7.1 (3H, m), 5.9 (1H, d), 4.2 (1H, m), 2.85 (3H, s), 1.65 (4H, m), 0.95 (6H, t); UV (EtOH):  $\lambda_{max} = 221$ , 241, 268, 298, 313 nm. Anal.  $C_{16}H_{20}N_4$  (C, H, N): calc. C 71.61, H 7.51, N 20.83%; found C 71.12, H 7.53, N 20.79%.

## 6.1.23. 8-Chloro-4-cyclopentylamino-1-methylimidazo [1,2-a]quinoxaline (24)

Recryst. solvent: petroleum ether/ethyl acetate. M.p. (DSC onset) 130.5 °C; IR (KBr): 3421, 2954, 1555 cm<sup>-1</sup>; 

<sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  7.95 (1H, d, J = 2 Hz), 7.6 (1H, d, J = 9 Hz), 7.35–7.15 (2H, m), 6.1 (1H, d), 4.5 (1H, m), 2.8 (3H, s), 2.4–2.05 (2H, m), 2.0–1.3 (6H, m); UV (EtOH):  $\lambda_{\text{max}}$  = 227, 251, 276, 305, 323, 336 nm. Anal. C<sub>16</sub>H<sub>17</sub>ClN<sub>4</sub> (C, H, N): calc. C 63.89, H 5.70, N 18.63%; found C 63.79, H 5.79, N 18.58%.

## 6.1.24. 8-Chloro-4-cyclohexylamino-1-methylimidazo [1,2-a]quinoxaline (25)

Recryst. solvent: *n*-hexane. M.p. (DSC onset) 130.4 °C; IR (KBr): 3410, 2927, 1551 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  7.95 (1H, d, J = 2 Hz), 7.6 (1H, d, J = 9 Hz), 7.4–7.2 (2H, m), 6.1 (1H, d), 4.2 (1H, m), 2.8 (3H, s), 2.3–1.1 (10H, m); UV (EtOH):  $\lambda_{\text{max}} = 227, 250, 276, 305, 323, 336$  nm. Anal.  $C_{17}H_{19}\text{CIN}_4$  (C, H, N): calc. C 64.86, H 6.08, N 17.80%; found C 64.86, H 6.21, N 17.91%.

## 6.1.25. 4-Cyclopentylamino-7,8-dichloro-1-methylimidazo[1,2-a]quinoxaline (26)

Recryst. solvent: *n*-hexane/ethyl acetate. M.p. (DSC onset) 213.5 °C; IR (KBr): 3411, 2960, 1548 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  8.0 (1H, s), 7.75 (1H, s), 7.25 (1H, s), 6.2 (1H, d), 4.6 (1H, m), 2.8 (3H, s), 2.3–1.3 (8H, m); UV (EtOH):  $\lambda_{max} = 233$ , 279, 308, 327, 341 nm. Anal.  $C_{16}H_{16}Cl_2N_4$  (C, H, N): calc. C 57.33, H 4.81, N 16.71%; found C 57.41, H 4.82, N 16.68%.

6.1.26. 8-Chloro-4-(1-ethylpropylamino)imidazo[1,2-a]quinoxaline (16)

A mixture of compound 29b (2.66 g, 12 mmol), hexamethyldisilazane (8.9 mL, 42 mmol), ammonium sulfate (0.32 g, 2.4 mmol) and 1-ethylpropylamine (7.0 mL, 60 mmol) was stirred at 120 °C in a Dean-Stark apparatus for 18 h. Upon cooling, the mixture was concentrated in vacuo to give a residue which was taken up in ethyl acetate plus water. The biphasic mixture contained a solid (consisting mainly of unreacted imidazoquinoxalinone) which was filtered away; the filtrate was collected and the organic layer was washed with brine, dried and evaporated, thus obtaining the crude title compound (0.97 g). Column chromatography (eluant: CH<sub>2</sub>Cl<sub>2</sub>) and recrystallization from n-hexane afforded pure 16. M.p. (DSC onset) 125.1 °C; IR (KBr): 3406, 3105, 2964, 1532 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>/CD<sub>3</sub>OD):  $\delta$  8.4 (1H, s), 8.05 (1H, d, J = 2 Hz), 7.8–7.4 (3H, m), 3.85 (1H, m), 1.65 (4H, m), 0.95 (6H, t); UV (EtOH):  $\lambda_{\text{max}} = 230$ , 245, 289, 301, 326 nm. Anal. C<sub>15</sub>H<sub>17</sub>ClN<sub>4</sub> (C, H, N): calc. C 62.39, H 5.93, N 19.40%; found C 62.17, H 6.02, N 19.46%.

The following compounds were similarly prepared from 1-methylimidazo[1,2-a]quinoxalin-4(5H)-one (30a) and the appropriate amine.

## 6.1.27. 4-Cyclopentylamino-1-methylimidazo[1,2-a] quinoxaline (20)

Recryst. solvent: ethyl acetate. M.p. (DSC onset) 167.3 °C; IR (KBr): 3294, 2948, 1544 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  8.0 (1H, dd), 7.7 (1H, dd), 7.4–7.05 (3H, m), 6.0 (1H, d), 4.6 (1H, m), 2.8 (3H, s), 2.3–2.05 (2H, m), 2.0–1.4 (6H, m); UV (EtOH):  $\lambda_{\text{max}} = 225$ , 243, 272, 301, 316, 329 nm. Anal. C<sub>16</sub>H<sub>18</sub>N<sub>4</sub> (C, H, N): calc. C 72.15, H 6.81, N 21.03%; found C 71.86, H 6.81, N 20.79%.

## 6.1.28. 4-Cyclohexylamino-1-methylimidazo[1,2-a] quinoxaline (21)

Recryst. solvent: *n*-hexane/ethyl acetate. M.p. (DSC onset) 126.5 °C; IR (KBr): 3345, 2938, 1546 cm<sup>-1</sup>; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  8.0 (1H, dd), 7.7 (1H, dd), 7.5–7.1 (3H, m), 6.0 (1H, d), 4.2 (1H, m), 2.85 (3H, s), 2.3–1.95 (2H, m), 1.95–0.9 (8H, m); UV (EtOH):  $\lambda_{\text{max}}$  = 225, 243, 272, 301, 316, 329 nm. Anal. C<sub>17</sub>H<sub>20</sub>N<sub>4</sub> (C, H, N): calc. C 72.83, H 7.19, N 19.98%; found C 72.21, H 7.54, N 20.14%.

## 6.1.29. General procedure for the synthesis of 2,3,6,7-tetrachloro- (31c), 2,3-dichloro-6-fluoro- (31d) and 2,3-dichloro-6,7-difluoroquinoxaline (31e)

The title compounds were prepared according to literature methods [21] from the appropriate 1,2-phenylenediamine, which in turn was commercially available (4,5-dichloro-1,2-phenylenediamine) or was

obtained by classical SnCl<sub>2</sub>/HCl reduction of the corresponding 2-nitroaniline. Briefly, a mixture of the phenylenediamine (10 mmol) and diethyl oxalate (60 mmol) was refluxed for 1 day. Upon cooling a solid precipitated; this was collected, washed with cold ethanol and dried to obtain the (substituted)-2,3-dihydroxyquinoxaline (yields 85–95%) which was subsequently treated with 5-fold molar excess of POCl<sub>3</sub>. The mixture was refluxed for 16 h; evaporation of the excess POCl<sub>3</sub> at reduced pressure afforded a residue which was taken up with CHCl<sub>3</sub>. The resulting suspension was filtered and the filtrate was washed with water and brine, then dried and evaporated to dryness to obtain the title compounds 31c–e (yields 75–90%).

#### 6.2. Biochemistry

Compounds were assessed for their ability to inhibit binding of the A<sub>1</sub>AR selective antagonist radioligand [3H]DPCPX to synaptosomal membranes from rat brain according to a published procedure [22]. Briefly, 200 µg of membrane proteins were incubated with test compound solution (at least ten different concentrations) and 0.3 nM [3H]DPCPX in 400 µL of 50 mM Tris.HCl, pH 7.4 for 1 h at 25 °C. Nonspecific binding was determined in the presence of 10  $\mu M$  (R)-N<sup>6</sup>-phenylisopropyladenosine (R-PIA). The incubations were blocked by filtration using a cell harvester and, upon separation of the bound from the free, the radioactivity contents were measured by liquid scintillation. All the assays were performed in triplicate or in quadruplicate. Data were fitted by nonlinear regression analysis (Allfit) and the  $K_i$ values were calculated according to the Cheng-Prusoff equation [33].

Compound **20** was assessed for its ability to inhibit binding of the  $A_{2a}AR$  selective agonist radioligand [ ${}^{3}H$ ]CGS21680 to rat striatal membranes according to a published procedure [23]. Briefly, 200 µg of membrane proteins were incubated with the test compound solution (twelve different concentrations) and 5 nM [ ${}^{3}H$ ]CGS21680 in the incubation buffer (50 mM Tris.HCl pH 7.4) for 1 h at 25 °C. Nonspecific binding was determined in the presence of 25 µM N<sup>6</sup>-cyclopentyladenosine (CPA). Subsequent harvesting of the assay and analysis of the data were identical with the  $A_1$  assay.

The primary screen for the evaluation of  $A_3AR$  binding affinity of compound **20** was performed by MDS Panlabs, Taipei, ROC, according to standardized procedures, using human recombinant HEK-293 cells [25] as the source, 0.4 nM [ $^{125}$ I]AB-MECA [24] as radioligand, and 10  $\mu$ M 5'-(*N*-ethylcarbamoyl)adenosine (NECA) for nonspecific binding.

#### Acknowledgements

We are grateful to Angela Tommasi, M. Letizia Marcoccia, Francesco Magnante and Rossella Piroli for their technical support in the chemical syntheses, as well as to dr. L. Dorigotti for his encouragement. We also thank prof. T. Boschi and his co-workers from the 2nd University of Rome 'Tor Vergata' who ran the 400 MHz <sup>1</sup>H-NMR spectra and performed the NOE-ROESY experiments.

#### References

- [1] Belardinelli L., Pelleg A. (Eds.), Adenosine and Adenine Nucleotides: From Molecular Biology to Integrative Physiology, Kluwer, Boston, 1995.
- [2] Olah M.E., Stiles G.L., Annu. Rev. Pharmacol. Toxicol. 35 (1995) 581–606.
- [3] Feoktistov I., Polosa R., Holgate S.T., Biaggioni I., Trends Pharmacol. Sci. 19 (1998) 148–153.
  - [4] Müller C.E., Stein B., Curr. Pharm. Des. 2 (1996) 501-530.
- [5] Van der Wenden E.M., Carnielli M., Roelen H.C., Lorenzen A., von Frijtag Drabbe Künzel J.K., Ijzerman A.P., J. Med. Chem. 41 (1998) 102–108.
- [6] Jakobson K.A., Kim H.O., Siddiqi S.M., Olah M.E., Stiles G.L., von Lubitz D.K.J.E., Drugs Fut. 20 (1995) 689-699.
- [7] Schingnitz G., Küfner-Mühl U., Ensinger H., Lehr E., Kuhn F.J., Nucleosides Nucleotides 10 (1991) 1067–1076.
- [8] Aki Y., Tomohiro A., Nishiyama A., Kiyomoto K., Kimura S., Abe Y., Pharmacology 55 (1997) 193–201.
- [9] Neely C.F., Jin J., Keith I.M., Am. J. Physiol. [Lung Cell. Mol. Physiol. 16] 272 (1997) L353-L361.
- [10] Sidi A., Wesley R., Barrett R., Rush W., Belardinelli L., Cardiovasc. Res. 28 (1994) 621–628.
  - [11] Mader T.J., Gibson P., Resuscitation 35 (1997) 3-7.
- [12] Jacobson K.A., van Galen P.J.M., Williams M., J. Med. Chem. 35 (1992) 407-422.
- [13] Pfister J.R., Belardinelli L., Lee G., Lum R.T., Milner P., Stanley W.C., Linden J., Baker S.P., Schreiner G., J. Med. Chem. 40 (1997) 1773–1778.
- [14] Van Galen P.J.M., Stiles G.L., Michaels G., Jacobson K.A., Med. Res. Rev. 12 (1992) 423–471.
- [15] Ceccarelli S., Altobelli M., D'Alessandro A., Paesano A., Res. Commun. Mol. Pathol. Pharmacol. 87 (1995) 101-102.
- [16] Francis J.E., Cash W.D., Psychoyos S., Ghai G., Wenk P., Friedmann R.C., Atkins C., Warren V., Furness P., Hyun J.L., Stone G.A., Desai M., Williams M., J. Med. Chem. 31 (1988) 1014–1020.
  - [17] Trivedi B.K., Bruns R.F., J. Med. Chem. 31 (1988) 1011-1014.
- [18] Van Galen P.J.M., Nissen P., van Wijngaarden I., Ijzerman A.P., Soudijn W., J. Med. Chem. 34 (1991) 1202–1206.
- [19] Sarges R., Howard H.R., Browne R.G., Lebel L.A., Seymour P.A., Koe B.K., J. Med. Chem. 33 (1990) 2240–2254.
- [20] Vorbrüggen H., Krolikiewicz K., Chem. Ber. 117 (1984) 1523–1541.
  - [21] Cheeseman G.W.H., J. Chem. Soc. (1962) 1170-1176.

- [22] Lohse M.J., Klotz K.-N., Lindenborn-Fotinos J., Reddington M., Schwabe U., Olsson R.A., Naunyn-Schmied. Arch. Pharmacol. 336 (1987) 204–210.
- [23] Jarvis M.F., Schulz R., Hutchison A.J., Do U.H., Sills M.A., Williams M., J. Pharmacol. Exp. Ther. 251 (1989) 888–893.
- [24] Olah M.E., Gallo-Rodriguez C., Jacobson K.A., Stiles G.L., Mol. Pharmacol.45 (1994) 978–982.
- [25] Salvatore C.A., Jacobson M.A., Taylor H.E., Linden J., Johnson R.G., Proc. Natl. Acad. Sci. USA 90 (1993) 10365–10369.
- [26] Dooley M.J., Kono M., Suzuki F., Bioorg. Med. Chem. 4 (1996) 923–934.
- [27] Peet N.P., Lentz N.L., Meng E.C., Dudley M.W., Ogden A.M.L., Demeter D.A., Weintraub H.J.R., Bey P.A., J. Med. Chem. 33 (1990) 3127–3130.

- [28] Ceccarelli S., D'Alessandro A., Magnante F., Scuri R., Zanarella S., Farmaco 48 (1993) 1301-1312.
- [29] Colotta V., Cecchi L., Catarzi D., Filacchioni G., Martini C., Tacchi P., Lucacchini A., Eur. J. Med. Chem. 30 (1995) 133-139.
- [30] Ceccarelli S., Altobelli M., D'Alessandro A., Dorigotti L., Drug Dev. Res. 43 (1998) 74 (P285).
- [31] Davey D.D., Erhardt P.W., Cantor E.H., Greenberg S.S., Ingebretsen W.R., Wiggings J., J. Med. Chem. 34 (1991) 2671-2677.
- [32] Mc Quaid L.A., Smith E.C.R., South K.K., Mitch C.H., Schoepp D.D., True R.A., Calligaro D.O., O'Malley P.J., Lodge D., Ornstein P.L., J. Med. Chem. 35 (1992) 3319–3324.
- [33] Cheng Y.C., Prusoff W.H., Biochem. Pharmacol. 22 (1973) 3099-3103.